

Lipid Nanocarriers as a Novel Platform for Precision Delivery of CRISPR-Based Gene Editing Systems and RNA Therapy in Glioblastoma

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ABSTRACT

Glioblastoma (GBM) is one of the most aggressive and lethal brain tumors, characterized by high heterogeneity, rapid progression, and resistance to current therapeutic strategies. Standard treatments—including surgery, radiotherapy, and temozolomide chemotherapy—have limited efficacy, and median survival remains approximately 15 months. Among various molecular drivers of GBM, microRNA-10b (miR-10b) plays a central oncogenic role, being consistently upregulated in glioma cells while absent in normal brain tissue. Its inhibition demonstrates strong therapeutic potential. However, a major challenge is the efficient and safe delivery of gene-editing systems to intracranial tumors. Recent advancements in lipid nanoparticles (LNPs) have made them a promising platform for delivering mRNA and sgRNA for CRISPR-based therapies.

Keywords: Glioblastoma, Lipid Nanoparticles, Gene Editing, CRISPR-Cas9, microRNA-10b

Introduction Glioblastoma (GBM) is one of the most aggressive primary brain tumors, with a median patient survival of approximately 15 months despite maximal surgical resection, radiotherapy and temozolomide (TMZ) chemotherapy (1,2). GBM treatment is hindered by extensive heterogeneity and the persistence of therapy-resistant glioma stem cells (GSCs) (3). MicroRNA-10b (miR-10b) has emerged as a uniquely important therapeutic target, due to its consistent upregulation in malignant gliomas, absence in normal brain tissue, and essential for glioma growth and survival (4–12). Although miR-10b gene editing has shown therapeutic potential, safe and efficient in vivo delivery of CRISPR–Cas9 systems to intracranial tumors remains a major challenge. **To overcome this challenge, researchers have developed lipid nanoparticles (LNPs) encapsulating Cas9 mRNA together with miR-10b–targeting sgRNAs, designated as miRTEN (13–16).**

Methods This review employed a structured literature analysis using recent peer-reviewed studies published in leading biomedical journals. Key databases including PubMed, Scopus, and Web of Science were systematically searched to identify advances in lipid nanoparticle design, CRISPR delivery strategies, and RNA-based therapeutics for glioblastoma.

Results Intracerebroventricular administration of miRTEN enabled broad and durable Cas9 mRNA expression and efficient miR-10b editing within both tumor cores and invasive

margins across multiple GBM models (5). miRTEN significantly reduced tumor growth, suppressed GSC proliferation, and increased tumor-cell death in a dose-dependent manner, with therapeutic effects correlating with the degree of miR-10b suppression (6). Combination therapy with TMZ further enhanced tumor inhibition, overcoming TMZ resistance and improving survival (17). In immunocompetent models, miRTEN activated robust antitumor immune responses, increased infiltration of cytotoxic CD8⁺ T cells, and induced durable immune memory capable of rejecting rechallenged tumors (18,19). Safety evaluations confirmed that miRTEN selectively targets GBM cells while sparing normal brain tissue, with no evidence of neurotoxicity or impaired neuronal activity (9).

Conclusion As CRISPR-based therapeutics advance toward clinical application, **researchers** findings demonstrate that LNP-mediated delivery of Cas9 mRNA and sgRNAs enables safe and effective miR-10b gene editing in vivo. miRTEN shows strong therapeutic benefit as both monotherapy and in combination with standard-of-care treatment, offering a promising platform for advancing gene-editing and RNA-based therapies for GBM and other high-grade gliomas.

References:

1. Mathur R, Wang Q, Schupp PG, et al. Glioblastoma evolution and heterogeneity from a 3D whole-tumor perspective. *Cell*. 2024;187(2):446–463.e16.
2. Schaff LR, Mellinghoff IK. Glioblastoma and other primary brain malignancies in adults: a review. *JAMA*. 2023;329(7):574–587.
3. Sharifzad F, Ghavami S, Verdi J, et al. Glioblastoma cancer stem cell biology: potential theranostic targets. *Drug Resist Updat*. 2019;42:35–45.
4. Teplyuk NM, Uhlmann EJ, Wong AH, et al. MicroRNA-10b inhibition reduces E2F1-mediated transcription and miR-15/16 activity in glioblastoma. *Oncotarget*. 2015;6(6):3770–3783.
5. Teplyuk NM, Uhlmann EJ, Gabriely G, et al. Therapeutic potential of targeting microRNA-10b in established intracranial glioblastoma: first steps toward the clinic. *EMBO Mol Med*. 2016;8(3):268–287.
6. Gabriely G, Yi M, Narayan RS, et al. Human glioma growth is controlled by microRNA-10b. *Cancer Res*. 2011;71(10):3563–3572.
7. El Fatimy R, Zhang Y, Deforzh E, et al. A nuclear function for an oncogenic microRNA as a modulator of snRNA and splicing. *Mol Cancer*. 2022;21(1):17.
8. Lin J, Teo S, Lam DH, Jeyaseelan K, Wang S. MicroRNA-10b pleiotropically regulates invasion, angiogenicity and apoptosis of tumor cells resembling mesenchymal subtype of glioblastoma multiforme. *Cell Death Dis*. 2012;3(10):e398–e398.
9. Guessous F, Alvarado-Velez M, Marcinkiewicz L, et al. Oncogenic effects of miR-10b in glioblastoma stem cells. *J Neurooncol*. 2013;112(2):153–163.
10. Ma C, Wei F, Xia H, et al. MicroRNA-10b mediates TGF- β 1-regulated glioblastoma proliferation, migration and epithelial-mesenchymal transition. *Int J Oncol*. 2017;50(5):1739–1748.
11. Sun B, Zhao X, Ming J, et al. Stepwise detection and evaluation reveal miR-10b and miR-222 as a remarkable prognostic pair for glioblastoma. *Oncogene*. 2019;38(33):6142–6157.

- 12.El Fatimy R, Subramanian S, Uhlmann EJ, Krichevsky AM. Genome editing reveals glioblastoma addiction to MicroRNA-10b. *Mol Ther.* 2017;25(2):368–378.
- 13.Finn JD, Smith AR, Patel MC, et al. A single administration of CRISPR/Cas9 lipid nanoparticles achieves robust and persistent in vivo genome editing. *Cell Rep.* 2018;22(9):2227–2235.
- 14.Eygeris Y, Henderson MI, Curtis AG, et al. Preformed vesicle approach to LNP Manufacturing Enhances Retinal mRNA Delivery. *Small* 2024;20(37):e2400815.
- 15.Gautam M, Jozic A, Su GL, et al. Lipid nanoparticles with PEG-variant surface modifications mediate genome editing in the mouse retina. *Nat Commun.* 2023;14(1):6468.
- 16.Couture-Senecal J, Natraj J, Khan OF. A cell-free kinetic analysis of ionizable lipid hydrolysis. *Anal Chem.* 2024;96(43):17128–17134.
- 17.Chen M, Kim B, Robertson N, et al. Co-administration of temozolomide (TMZ) and the experimental therapeutic targeting miR-10b, profoundly affects the tumorigenic phenotype of human glioblastoma cells. *Front Mol Biosci.* 2023;10:1179343.
- 18.Tsukerman P, Stern-Ginossar N, Gur C, et al. MiR-10b downregulates the stress-induced cell surface molecule MICB, a critical ligand for cancer cell recognition by natural killer cells. *Cancer Res.* 2012;72(21):5463–5472.
- 19.Tsukerman P, Enk J, Mandelboim O. Metastamir-mediated immune evasion: miR-10b downregulates the stress-induced molecule MICB, hence avoid recognition by NKG2D receptor. *Oncoimmunology* 2013;2(1):e22245.